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# **itero Documentation**

*Release 1.1*

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Release v1.1. (*Changelog*)

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*itero* is a “guided-assembly” workflow for target enrichment data that uses *bwa*, *samtools*, *bedtools*, and *spades* to provided high-quality assemblies of raw reads from *Illumina* instruments.



# CHAPTER 1

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## Contributions

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`itero` is open-source (see [License](#)) and we welcome contributions from anyone who is interested. Please make a pull request on [github](#). The issue tracker for `itero` is also available on [github](#).



## CHAPTER 2

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### Issues

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If you have an issue, please ensure that you are experiencing this issue on a supported OS (see *Installation*) using the `conda` installation of `itero`. If possible, please submit a test case demonstrating the issue and indicate which platform, git checkout, and phyluce version you are using.



## 3.1 Purpose

`itero` is a program that wraps a workflow for guided- or reference-based assembly of target enrichment data. This approach to assembly is also “iterative”, meaning that the assembly proceeds through several, iterations (hence “itero”). I wrote `itero` for a variety of reasons:

- “traditional” DNA assembly programs performed poorly with target enrichment data (from UCE loci)
- existing DNA assembly approaches had relatively high assembly error
- existing guided assembly programs were hard to install and run
- some existing guided assembly programs were **slow**

`itero` attempts to fix some of these problems. At its heart, `itero` uses an input file of “seeds”, against which it will try to assemble raw-read data from **Illumina** instruments. Alignment of reads-to-seeds uses `bwa`, the BAM file is split with `samtools` and `bedtools`, and locus-specific reads are then assembled using `spades` (with error correction turned on during the final round). Then, the entire process repeats itself.

To increase assembly speed, `itero` takes advantage of multiple cores (on single nodes) using `python` multiprocessing and MPI (on HPC systems) using the excellent `schwimmbad` library.

### 3.1.1 Who wrote this?

This documentation was written primarily by Brant Faircloth (<http://faircloth-lab.org>). Brant is also responsible for the development of most of the `itero` code. Bugs within the code are usually his.

### 3.1.2 How do I report bugs?

To report a bug, please post an issue to <https://github.com/faircloth-lab/itero/issues>. Please also ensure that you are using one of the “supported” platforms:

- Apple OSX 10.9.x

- CentOS 7.x

and that you have installed `itertools` and dependencies using `conda`, as described in the *Installation* section.

## 3.2 Installation

`itertools` uses a number of `Python` tools that allow it to assemble raw reads into contigs. `itertools` also wraps a number of third-party programs. These include:

### 3.2.1 Python Modules

- `numpy`
- `biopython`
- `mpi4py`
- `schwimmbad`
- `six`

### 3.2.2 3rd-party programs

- `bedtools`
- `bwa`
- `gawk`
- `samtools`
- `spades`

To ensure that these dependencies are easy to install, we have created a `conda` package for `itertools` that is available as part of `bioconda`. This is the easiest way to get `itertools` up and running on your system. `itertools` can also be run outside of `conda`, and we include some installation suggestions for these types of systems, below. However, because many HPC systems are configured differently, we cannot provide extensive support for `itertools` on HPC platforms.

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**Note:** We build and test the binaries available through `conda` using 64-bit operating systems that include:

- Apple OSX 10.9.x
  - CentOS 7.x
- 

### 3.2.3 Why conda?

It may seem odd to impose a particular distribution on users, and we largely agree. However, `conda` makes it very easy for us to distribute both `Python` and non-`Python` packages, setup identical environments across very heterogeneous platforms (linux, osx), make sure all the `$PATHs` are correct, and have things run largely as expected. Using `conda` has several other benefits, including environment separation similar to `virtualenv`.

In short, using `conda` gets us as close to a “one-click” install that we will probably ever get.

### 3.2.4 Install Process (using conda/bioconda)

**Attention:** We do not support `itero` on Windows.

**Note:** We build and test the binaries available through using 64-bit operating systems that include:

- Apple OSX 10.9.x
- CentOS 7.x

The installation process is a 3-step process. You need to:

1. Install `conda` (either `anaconda` or `miniconda`)
2. Configure `conda` to use `bioconda`
3. Install `itero`

Installing `itero` using `conda` will install all of the required binaries, libraries, and `Python` dependencies.

#### Install Anaconda or miniconda

You first need to install `anaconda` or `miniconda`. Which one you choose is up to you, your needs, how much disk space you have, and if you are on a fast/slow connection.

**Attention:** You can easily install `anaconda` or `miniconda` in your `$HOME`, although you should be aware that this setup can cause problems in some HPC setups.

**Tip:** Do I want `anaconda` or `miniconda`?

The major difference between the two python distributions is that `anaconda` comes with many, many packages pre-installed, while `miniconda` comes with almost zero packages pre-installed. As such, the beginning `anaconda` distribution is roughly 500 MB in size while the beginning `miniconda` distribution is 15-30 MB in size.

#### `anaconda`

Follow the instructions here for your platform: <http://docs.continuum.io/anaconda/install.html>

#### `miniconda`

Find the correct `miniconda-x.x.x` file for your platform from <http://repo.continuum.io/miniconda/> and download that file. Be sure you **do not** get one of the packages that has a name starting with `miniconda3-`. When that has completed, run one of the following:

```
bash Miniconda-x.x.x-Linux-x86_64.sh [linux]
bash Miniconda-x.x.x-MacOSX-x86_64.sh [osx]
```

**Note:** Once you have installed Miniconda, we will refer to it as **anaconda** throughout the remainder of this documentation.

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### Checking your `$PATH`

Regardless of whether you install `anaconda` or `miniconda`, you need to check that you've installed the package correctly. To ensure that the correct location for `anaconda` or `miniconda` are added to your `$PATH` (this occurs automatically on the `$BASH` shell), run the following:

```
$ python -V
```

The output should look similar to (*x* will be replaced by a version):

```
Python 2.7.x :: Anaconda x.x.x (x86_64)
```

Notice that the output shows we're using the *Anaconda x.x.x* version of `Python`. If you do not see the expected output (or something similar), then you likely need to edit your `$PATH` variable to add `anaconda` or `miniconda`.

The easiest way to edit your path, if needed is to open `~/ .bashrc` with a text editor (if you are using `ZSH`, this will be `~/ .zshrc`) and add, as the last line:

```
export PATH=$HOME/path/to/conda/bin:$PATH
```

where `$HOME/path/to/conda/bin` is the location of `anaconda/miniconda` on your system (usually `$HOME/anaconda/bin` or `$HOME/miniconda/bin`).

**Warning:** If you have previously set your `$PYTHONPATH` elsewhere in your configuration, it may cause problems with your `anaconda` or `miniconda` installation of `phyluce`. The solution is to remove the offending library (-ies) from your `$PYTHONPATH`.

### Configure Bioconda

Once you have installed `anaconda` (or `miniconda`), you need to configure `conda` to use the `bioconda` channel. More information on this process can be found on the `bioconda` website, but the gist of the process is that you need to run:

```
conda config --add channels defaults
conda config --add channels conda-forge
conda config --add channels bioconda
```

### Install itero

Once `bioconda` is installed, you should be able to install `itero` by running:

```
conda install itero
```

This will install everything that you need to run the program.

### Test itero install

You can check to make sure all of the binaries are installed correctly by running:

```
itero check binaries
```

### 3.2.5 Install Process (Alternative / HPC)

On some systems (particularly HPC systems), `conda` can cause problems. You can `itero` the “old” way by downloading the package tarball (<https://github.com/faircloth-lab/itero/releases>) and running:

```
python setup.py install
```

in the main directory. This should install all of the `Python` dependencies, **but you still need to install and configure the 3rd-party dependencies.**

**Attention:** You will need to install 3rd-party dependencies on your own if you are using the `python setup.py install` method of installing `itero`

You can build and install these dependencies where you like. To configure `itero` to use the dependencies you have build and installed, you need to create a `$HOME/.itero.conf` that gives the paths to each program and looks like:

```
[executables]
bedtools:/path/to/bin/bedtools
bwa:/path/to/bin/bwa
gawk:/path/to/bin/gawk
samtools:/path/to/bin/samtools
spades:/path/to/bin/spades.py
```

### Test itero install

You can check to make sure all of the binaries are installed correctly by running:

```
itero check binaries
```

## 3.3 Running itero

`itero` has both a **local** mode and an **MPI** mode. The **local** mode is for execution on a single node, while the **MPI** mode executes individual locus assemblies in parallel using an MPI-enabled HPC cluster. To run the program, you must first create a configuration file denoting the samples you wish you assemble. That file has the following format:

```
[reference]
/path/to/the/locus/seeds.fasta

[individuals]
taxon-one:/path/to/fastq/R1/and/R2/files/for/taxon/1/
taxon-two:/path/to/fastq/R1/and/R2/files/for/taxon/2/
taxon-three:/path/to/fastq/R1/and/R2/files/for/taxon/3/
```

### 3.3.1 itero on a single node

You then run the *local* version using a command similar to:

```
itero assemble local --config ndna-test.conf
--output local
--local-cores 16
--iterations 6
```

This will run *itero* on a single node and will first use 16 cores to perform *bwa* alignments. The code will then distribute locus-specific assemblies across all cores on the node (1 assembly per core; 16 in parallel).

### 3.3.2 itero across MPI nodes

You run the *MPI* version using a command similar to:

```
mpirun -hostfile hostfile -n 96 itero assemble mpi --config ndna-test.conf \
--output mpi \
--local-cores 16 \
--iterations 6
```

If each of your nodes has 16 cores, this will first use 16 cores for the needed *bwa* alignments of reads to seeds. The code will then distribute locus-specific assemblies across all 96 cores in your cluster (1 assembly per core; 96 in parallel).

## 4.1 License

### 4.1.1 Documentation

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If you use this documentation or the `itero` software for your own research, please cite both the software and (Faircloth et al. 2012). See the Citing section for more detail.

### 4.1.2 Software

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## 4.2 Changelog

### 4.2.1 v1.0.x (April 2018)

- initial version with MPI and multiprocessing capability

### 4.2.2 v1.1.0 (May 2018)

- fix error in contig checking code that could cause MPI operations to hang
- refactor BAM splitting code for hopefully faster operation
- add RAM limits on spades
- add configuration parameters to iter.conf for spades
- create unique log file for each run

### 4.2.3 v1.1.1 (June 2018)

- fix an error where too many fastq files would cause MPI to hang

## 4.3 Funding

### 4.3.1 Primary Sources

The [National Science Foundation \(NSF\)](#) has supported a large portion of our work. The specific programs and proposal identifiers are below:

- NSF DEB-1655624
- NSF IOS-1754417